

INTRODUCTION

Tumour Growth Factor β (TGF β) isoforms belong to a super-family of cytokines that regulate cell growth and differentiation, cell motility, angiogenesis and apoptosis. Signal is transduced by a family of serine-threonine kinase receptors through activation of SMAD proteins. In cancer, activation primarily occurs after the formation of a complex between TGF β R1 and TGF β R2.

Previous *in vitro* studies have suggested that microglia may promote tumour growth via production of TGF β family members in the tumour microenvironment.

Our aim is to investigate the expression of TGF β and TGF β R in astrocytomas and oligodendrogliomas, identify its cell source and discuss its possible role as therapeutic target in patients with.

METHOD

We first reviewed 6 microarray gene profile datasets available in GEO (<http://www.ncbi.nlm.nih.gov/sites/entrez>) assessing for expression of TGF β I and II in human gliomas.

Fifty supratentorial adult gliomas (10 diffuse astrocytoma, 10 anaplastic astrocytoma, 20 glioblastoma, 4 low grade and 6 anaplastic oligodendrogliomas) were retrieved from the BTRC Tumour Registry at the Charing Cross Hospital. Patients ages ranged between 25 and 68 years; all received surgical debulking. High grade tumours were then treated with radiotherapy and chemotherapy.

TGF β 1 and TGF β 2 expression and the extent of microglial infiltration were assessed on tissue sections by immunohistochemistry using immunoperoxidase and double immunofluorescence. Antibodies used are shown in Table 1.

TABLE 1

	Manufacturer	Clone	Dilution	Pre-treatment
TGF β I	Santa Cruz Biotechnology	Poly-clonal (TB21)	1:1200	Citrate buffer + 30min MW
TGF β II	Santa Cruz Biotechnology	Poly-clonal (V)	1:400	None
TGF β RII	Santa Cruz Biotechnology	Poly-clonal (V)	1:200	None
Iba1	Santa Cruz Biotechnology	Mono-clonal (1022-5)	1:75	EDTA + 40min MW

RESULTS

Dataset analysis: The microarray dataset expression of TGF β genes showed no significant correlation between relative gene expression and glioma grade, either between datasets or within separate probes themselves in each individual dataset.

Immunohistochemistry: In all cases, TGF β I expression was restricted to tumour endothelium and perivascular macrophages. TGF β II expression localised to tumour cells in a subset of 9 astrocytomas (1 low grade astrocytoma, 2 anaplastic astrocytomas and 6 glioblastomas) (Fig. 1). No TGF β I was observed in oligodendrogliomas and no expression was seen in microglial cell (Fig. 2).

Double immunofluorescence studies using Iba-1 confirmed the lack of TGF β II in microglia across all tumour subtypes (Fig. 2).

The immunoreaction for TGF β RII was positive in endothelial cells. A non-specific nuclear immunolabelling was seen in neoplastic cells but no obvious expression was observed.

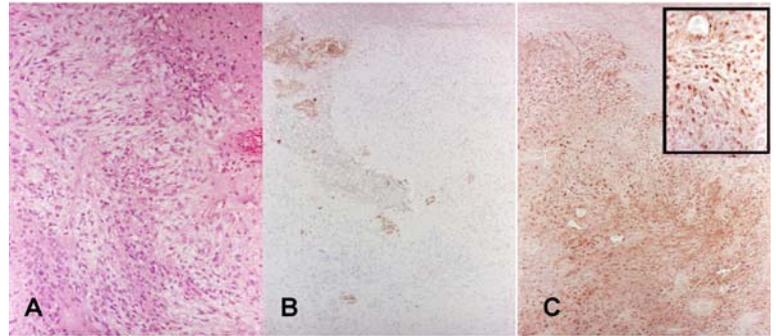


Fig 1: This case of glioblastoma (A, HE 10x) shows no TGF β I expression in tumour cells (B, perox 10x) whereas TGF β II is extensively positive (C, perox 10x – inset 40x)

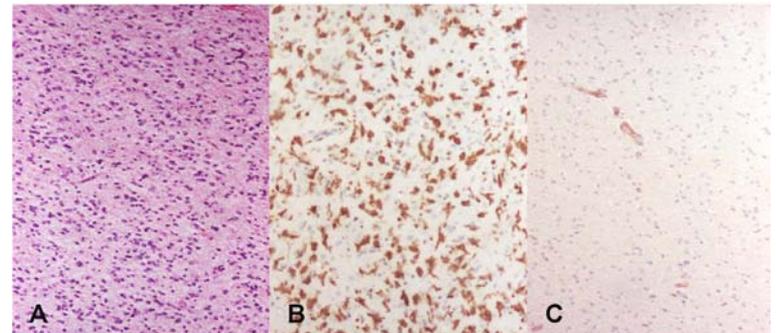


Fig 2: This example of anaplastic astrocytoma (A, HE 20x) show considerable microglial activation (B, Iba1 perox 20x); no TGF β II is conversely present in tumour and microglial cells (C, perox 10x)

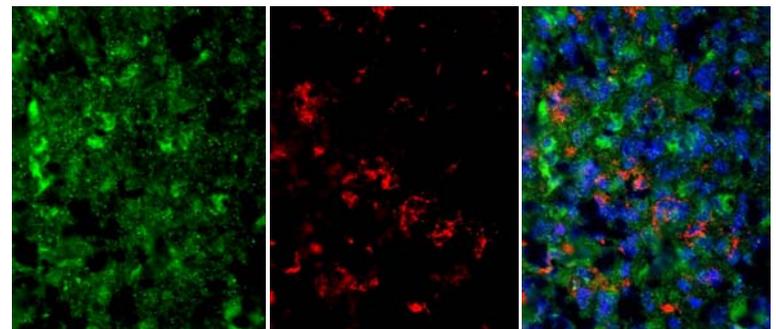


Fig 2: The colocalisation of Iba1 (A, green, 20x) and TGF β II (B, red, 20x) confirms the lack of TGF β II expression in microglial cells (C, merge – nuclear DAPI, 20x)

CONCLUDING REMARKS

Our results identify a subgroup of astrocytomas expressing TGF β II and confirm the variability of expression seen with the analysis of expression microarrays.

No autocrine/paracrine mechanism between neoplastic cells and microglial cells is observed. We are therefore unable to confirm *in vitro* studies suggesting that microglial-derived TGF β I promotes glioma growth and invasion *in vivo*.

Our results do not presently support the use of TGF β R antagonist in the treatment of human gliomas.

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